Accurate measurement of ¹³C-¹⁵N distances with solid-state NMR

Jae-Seung Lee and A. K. Khitrin^{a)}

Department of Chemistry, Kent State University, Kent, Ohio 44242-0001

(Received 17 February 2006; accepted 7 March 2006; published online 14 April 2006)

Solid-state NMR technique for measuring distances between heteronuclei in static powder samples is described. It is based on a two-dimensional single-echo scheme enhanced with adiabatic cross polarization. As an example, the results for intramolecular distances in α -crystalline form of glycine are presented. The measured NMR distances ${}^{13}C_{\alpha} - {}^{15}N$ and ${}^{13}C' - {}^{15}N$ are 1.496±0.002 and 2.50±0.02 Å, respectively. © 2006 American Institute of Physics. [DOI: 10.1063/1.2191043]

I. INTRODUCTION

Nuclear magnetic resonance (NMR) is a powerful tool for studying molecular structures. Information about internuclear distances is retrieved from magnitudes of long-range dipole-dipole interactions. In liquids, where dipolar couplings are averaged out by fast molecular motions, interatomic distances can be estimated from nuclear Overhauser effect.¹ More quantitative information is obtained with using anisotropic liquid-crystalline solvents,² which partially restore the dipole-dipole couplings. Orientation with bicelles became an efficient technique to determine structures of complex biomolecules.³ For molecules of liquid crystal itself, dipolar couplings provide valuable information about structure and ordering.^{4–6}

More accurate data on internuclear distances can be obtained for solids, where the effect of molecular motions is small. Various techniques, such as separated-local-field spectroscopy⁷ or polarization inversion spin exchange at magic angle (PISEMA),^{8,9} have been developed for measuring distances between rare and abundant nuclei. Since quality of heteronuclear decoupling can be substantially higher than that of homonuclear decoupling, one would expect better accuracy for well-isolated pairs of rare nuclei when abundant nuclei (protons) are decoupled. Today, most of the solidstate NMR experiments use magic-angle spinning (MAS) of a sample, which, by eliminating chemical shift anisotropy in powder samples, solves two problems: allows site assignments and increases signal-to-noise (S/N) ratio by creating narrow spectral peaks. MAS also averages the dipoledipole couplings, and they should be reintroduced by using a rotational resonance.¹⁰ Rotor-synchronized recoupling sequences¹¹ make it possible to combine site assignments with information about dipolar couplings. When the pairs of rare nuclei are created by specific isotope labeling and, therefore, the problem of assignment is solved, the most popular techniques for measuring internuclear distances under MAS are rotational echo double resonance (REDOR) (Ref. 12) (for heteronuclear pairs) and buildup of the double-quantum coherence¹³ (for homonuclear pairs). In both of these techniques the experimental spin dynamics under MAS is compared to results of computer simulation. Such simulation requires additional information about tensors of chemical shifts and their orientations with respect to the internuclear vector. Typical accuracy of measuring the constants of dipole-dipole interaction under MAS is about 10%. More accurate NMR measurements have been performed for static samples. Among the techniques are the nutation spectroscopy¹⁴ or use of the Carr-Purcell sequence¹⁵ for eliminating the chemical shift interaction in homonuclear systems, and MLEV-8 sequence with composite pulses¹⁶ in heteronuclear systems. When continuous or multipulse irradiation is strong (but, in practice, not infinitely strong) the system evolves under the influence of an effective Hamiltonian, which is known only approximately. To solve this problem, in Ref. 15 the experimental data were extrapolated to infinite intervals between pulses in a Carr-Purcell sequence. However, there is a problem with such extrapolation for homonuclear systems: the frequency of dipolar oscillations of a two-spin system depends on a difference of chemical shifts. For the same orientation of the internuclear vector, say perpendicular to the magnetic field, different orientations of crystals, allowed by rotations around this internuclear vector, create a distribution of the chemical shift differences, if the chemical shift tensors are not exactly the same for both nuclei. The situation is much better for heteronuclear pairs when the dipole-dipole interaction is truncated to ZZ term and commutes with the difference of chemical shifts. Therefore, the extrapolation to infinite intervals between pulses for a heteronuclear system¹⁶ is conceptually correct. At the same time, in both cases of using the extrapolation to infinite intervals between pulses^{15,16} the difference between actually measured and extrapolated values exceeded 10%.

We believe that better accuracy can be achieved when perturbation of the system is physically reduced to minimum and most of the time the system evolves under the unperturbed Hamiltonian of a heteronuclear two-spin system,

$$H = \omega_I I_z + \omega_S S_z + \left[\frac{\mu_0}{4\pi} \frac{\gamma_I \gamma_S \hbar}{r^3} (1 - 3\cos^2\theta) + J \right] I_z S_z, \quad (1)$$

where $\omega_I(\omega_S)$ and $\gamma_I(\gamma_S)$ are the chemical shift and the gyromagnetic ratio of spin I(S), μ_0 is the permeability of free space, r is the distance between spins I and S, θ is the angle between the static magnetic field and the internuclear vector connecting spins I and S, and J is the isotropic spin-spin

0021-9606/2006/124(14)/144508/5/\$23.00

124, 144508-1

^{a)}Electronic mail: akhitrin@kent.edu



FIG. 1. Experimental pulse sequence consists of adiabatic cross polarization, evolution (t_1) , and signal acquisition (t_2) periods.

coupling constant. Refocusing of the chemical shift interactions can be performed with a single pair of simultaneous 180° pulses on both of the heteronuclei. Insufficient sensitivity may be the major difficulty in detecting static powder Pake doublets from spin pairs at low concentration. However, as we demonstrate below, a combination of adiabatic cross polarization¹⁷ and efficient heteronuclear decoupling^{18–21} makes it possible to study even strongly-coupled ¹³C-¹⁵N pairs at concentrations 1% or less.

II. SCHEME OF EXPERIMENT

The scheme of experiment is shown in Fig. 1. It resembles the pulse sequence used in two-dimensional heteronuclear *J* spectroscopy.^{22,23} Our scheme starts with adiabatic cross polarization¹⁷ (ACP) to boost polarization of the ¹³C nuclei. ACP consists of adiabatic demagnetization/ remagnetization in the laboratory frame (ADLF/ARLF), performed with two frequency-sweeping pulses. The efficiency of ACP is illustrated in Fig. 2, showing about sevenfold increase of the ¹³C polarization in glycine. T_1 times for ¹H and ¹³C are about 0.5 and 1.5 s, respectively. The repetition time of 25 s was used for acquiring both of the spectra in Fig. 2, and it has been checked that the results do not change with further increase of the repetition time.

After ACP, ¹H spins are decoupled, and a 90° pulse creates transverse magnetization of ¹³C spins. The spin system consisting of ¹³C and ¹⁵N spin pairs evolves under their dipole-dipole interaction, while the two simultaneous 180° pulses on both spins in the middle of the evolution period t_1 refocus dephasing by chemical shifts and form an echo at the end of the evolution period. The ¹³C NMR signal is acquired



FIG. 2. ¹³C NMR spectrum of 2% glycine-[${}^{13}C_{\alpha}$, ${}^{15}N$] in n. a. glycine: (a) thermal equilibrium spectrum; (b) the spectrum enhanced with adiabatic cross polarization.



FIG. 3. Dipolar spectra for (a) 1% glycine-[${}^{13}C_{\alpha}$, ${}^{15}N$] in glycine-[${}^{12}C_{2}$, ${}^{14}N$] (exp. time ~16.5 h); (a') 2% glycine-[${}^{13}C_{\alpha}$, ${}^{15}N$] in n. a. glycine (exp. time ~20.5 h); (b) 1% glycine-[${}^{13}C'$, ${}^{15}N$] in glycine-[${}^{12}C_{2}$, ${}^{14}N$] (exp. time ~20.5 h); (b') 2% glycine-[${}^{13}C'$, ${}^{15}N$] in n. a. glycine (exp. time ~20.5 h).

in a two-dimensional way by varying the evolution time. The Fourier transform with respect to the evolution time t_1 gives the ${}^{13}\text{C}{}^{-15}\text{N}$ dipolar powder spectrum.

III. SAMPLES AND EXPERIMENTAL PARAMETERS

To test this scheme we have used α -crystalline form of glycine which has been previously studied with various techniques including NMR,^{16,24} x ray,²⁵ and neutron scattering.^{26–28} Isotopically labeled glycine-[$^{13}C_{\alpha}$, ^{15}N] (ISO-TEC) has been used as a test for short ${}^{13}C - {}^{15}N$ distance, and glycine-[¹³C', ¹⁵N] (Aldrich) for longer ¹³C-¹⁵N distance. Labeled glycine was diluted to 2% in natural abundance (n.a.) glycine (Aldrich) and to 1% in isotopically depleted glycine-[¹²C₂, ¹⁴N] (ISOTEC). All four samples were recrystallized from aqueous solutions. The experiments have been performed at room temperature (25 °C) with a Varian Unity/ Inova 500 MHz NMR spectrometer using a triple resonance "indirect" probe for liquids. In the ACP period, the first ¹H adiabatic pulse had 100 ms duration, 100 kHz frequencysweeping range, and 1.2 kHz amplitude $(\gamma B_1/2\pi)$; the second ¹³C pulse had 100 ms duration, 60 kHz frequencysweeping range, and 9.5 kHz amplitude. In the evolution period, the 90° and 180° pulses on ¹³C spin were, respectively, 16 and 32 μ s long, and the 180° pulse on ¹⁵N spin was 80 μ s long. The number of t_1 increments was 128, and the longest echo time was 50.8 ms. Proton spins were decoupled during the evolution and acquisition periods by using the SPINAL-32 heteronuclear decoupling sequence.¹⁹ The signal acquisition time t_2 was set to 2 ms.

IV. RESULTS

Figure 3 shows the dipolar powder spectra obtained for (a) 1% glycine- $[{}^{13}C_{\alpha}, {}^{15}N]$ in isotopically depleted glycine- $[{}^{12}C_2, {}^{14}N]$, (a') 2% glycine- $[{}^{13}C_{\alpha}, {}^{15}N]$ in naturalabundance glycine, (b) 1% glycine- $[{}^{13}C', {}^{15}N]$ in glycine- $[{}^{12}C_2, {}^{14}N]$, and (b') 2% glycine- $[{}^{13}C', {}^{15}N]$ in naturalabundance glycine. Both the "singular" peaks contributed by

Downloaded 16 Apr 2006 to 131.123.233.187. Redistribution subject to AIP license or copyright, see http://jcp.aip.org/jcp/copyright.jsp



FIG. 4. Simulated spectra for different relative broadening factors: 2%, 5%, and 10%. Although the peak positions shift with increasing broadening factor, the spectral intervals between the points at 90% of relative peak intensity can be consistently measured, as shown in the inset.

the internuclear vectors perpendicular to the magnetic field $(\theta \approx \pi/2)$ and steplike boundaries, which come from the parallel orientation $(\theta \approx 0)$, are clearly seen in the dipolar powder spectra. The central peaks originate from ¹³C spins of the matrix and nonperfect 180° pulses (rf field inhomogeneity). The dipolar frequency $\omega_D = (\mu_0/4\pi)\gamma_I\gamma_5\hbar/r^3$ and, therefore, NMR distance $r_{\rm NMR} = ((\mu_0/4\pi)(\gamma_I\gamma_5\hbar/\omega_D))^{1/3}$ can be extracted from the spectra in Fig. 3 by using simple data processing described in the following section.

V. DATA PROCESSING

There are three major factors that create distortions of the spectra in Fig. 3 and make them differ from the ideal theoretical dipolar powder spectra: line broadening, which mostly comes from nonperfect proton decoupling in our experiments; scalar J coupling, which should be taken into account for the case of the nearest neighbors [Figs. 3(a) and 3(a')]; and the delayed acquisition in t_1 dimension, which is due to finite pulses duration. Intensity of the measured signal decays with increasing evolution time t_1 . This line broadening shifts the "singular" peaks at $\pm \omega_D/2$ toward the center of the spectrum. Under the assumption that the signal decays exponentially with t_1 , the spectrum is a convolution of the ideal dipolar powder spectrum and the Lorentzian shape. Simulated theoretical spectra for three different broadening factors are shown in Fig. 4. Even though the peak positions shift toward the center of the spectrum at increasing broadening factor, one can notice that all the spectra cross the frequency $\pm \omega_D/2$ at the same relative height, about 90% of the maximum (see the inset in Fig. 4). This suggests a simple method of correcting for the broadening factor: the spectral interval should be measured between the two outside points at 90% of relative intensity.

For the directly bonded ¹³C and ¹⁵N nuclei, the scalar J coupling should also be taken into account. Although the value of J coupling can be obtained from fitting the spectrum in Fig. 3(a), we preferred not to introduce an extra fitting parameter and used the value J=6.7 Hz measured separately in a solution. The signs of J coupling and the dipolar cou-

TABLE I. Experimental results for intramolecular ${}^{13}\text{C}-{}^{15}\text{N}$ distances in α -glycine. $(\omega'_D+J)/2\pi$ is the frequency interval at 90% height (see text), $4\Delta f$ is the measured line-broadening factor, and $\omega_D/2\pi$ is the dipolar coupling constant after correction for the acquisition delay. The data in parentheses are obtained with ${}^{15}\text{N}$ detection.

Sample	$(\omega_D'+J)/2\pi$ (Hz)	$4\Delta f$ (Hz)	$\omega_D/2\pi$ (Hz)	r _{NMR} (Å)
1% glycine- $[{}^{13}C_{\alpha}, {}^{15}N]$	929.21	35.9	915.21	1.4959
in glycine- $[^{12}C_2, ^{14}N]$	(924.64)	(14.8)	(914.29)	(1.4964)
2% glycine- $[{}^{13}C_{\alpha}, {}^{15}N]$	925.67	59.4	906.97	1.5004
in n. a. glycine	(923.40)	(17.9)	(912.28)	(1.4975)
1% glycine-[¹³ C', ¹⁵ N]	195.62	15.9	194.94	2.5048
in glycine- $[^{12}C_2, ^{14}N]$	(196.07)	(13.4)	(195.36)	(2.5029)
2% glycine-[¹³ C', ¹⁵ N]	197.80	25.8	196.68	2.4973
in n. a. glycine	(196.22)	(15.5)	(195.40)	(2.5028)

pling at $\theta = \pi/2$ are the same (negative). Therefore, 6.7 Hz was subtracted from the frequency interval between the points at 90% height.

The distortion from the pair of 180° pulses is small but not zero. Finite duration of the pulses and after-pulse delays creates an acquisition delay τ in the τ_1 dimension. In the absence of line broadening, the acquisition delay affects the shape of the spectrum but does not shift the positions of the singular peaks. Our numerical study showed that there exists interference between line broadening and acquisition delay, which shifts the peaks outside. The corresponding frequency shift is well described by the following empirical formula:

$$\frac{\Delta\omega}{2\pi} = 2.2 \frac{\omega_D}{2\pi} (4\Delta f)\tau,\tag{2}$$

where Δf is the distance in frequency between the points of 90% and 70% intensities on the outside slope of a "singular" peak. The Lorentzian broadening factor is $4\Delta f$.

The step-by-step results following this data processing scheme are shown in Table I. First, the distances between the points of 90% intensity are determined and adjusted by subtracting the spin-spin coupling constant J. This value is denoted ω'_D in the Table I. Then, the line broadening factors are estimated from the frequency interval between the points of 90% and 70% intensities on the outside slope of a "singular" peak. The additional shift, resulting from the acquisition delay, is calculated with Eq. (2) and subtracted from ω'_D to give $\omega_D = \omega'_D - \Delta \omega$. The internuclear distances $r_{\rm NMR}$ are calculated as $((\mu_0/4\pi)(\gamma_1\gamma_s\hbar/\omega_D))^{1/3}$.

A comparison of the spectrum in Fig. 3(a) with the theoretical one is presented in Fig. 5. In the theoretical spectrum, we used the experimental value of the acquisition delay τ =100 μ s, which is a sum of the ¹⁵N pulse duration (80 μ s) and two after-pulse delays (10 μ s each). Some deviations between the theoretical and experimental spectra are due to the fact that the line-broadening factor depends on crystals orientation and, as a result, the total spectrum cannot be fitted using a uniform line broadening. This is one of the reasons why we used narrow spectral regions of the outside slopes of the "singular" peaks for retrieving information about dipolar coupling constants, rather than trying to extract this information from fitting the total powder pattern.



FIG. 5. Comparison of experimental (thin line) and theoretical (thick line) powder spectra for 1% glycine-[$^{13}C_{\alpha}$, ^{15}N] in isotopically depleted glycine-[$^{12}C_2$, ^{14}N].

Fitting with variable ω_D and Δf of the outside slopes of the "singular" peaks of the spectrum in Fig. 3(a) was used to estimate a statistical error of the results. It showed very small uncertainties of ±1 Hz for ω_D and ±3 Hz for Δf , determined as deviations from the best fit values where the rms difference between the experimental and theoretical spectra doubles.

Correlation between chemical shifts and orientation of the internuclear vector, in the presence of line broadening, create small differences between traces of the twodimensional spectrum. The spectra shown in Fig. 3 are socalled "sky projections" obtained from the real (cosine) Fourier transforms with respect to the evolution time t_1 , using the Varian's VNMRJ software. The values of ω_D calculated from different traces are distributed in a range ±5 Hz. Since we did not analyze the correlation between chemical shifts and orientations of the internuclear vector, we treated this difference between traces as a source of error and estimated, conservatively, the relative errors in r_{NMR} to be about 0.15% and 0.8% for the short and long distances, respectively.

VI. ATOMIC VIBRATIONS

The dipolar coupling constant ω_D measured by NMR is a quantity averaged over molecular motions. Modulation of the distance *r* between nuclei and the angle θ affect the measured ω_D and, therefore, the calculated distance r_{NMR} . This effect can be clearly seen when one compares the distances measured by NMR with the ones obtained from x-ray or



FIG. 6. Dipolar powder spectra obtained with ¹⁵N detection: (a) 1% glycine-[¹³C_a, ¹⁵N] in glycine-[¹²C₂, ¹⁴N] (exp. time ~80 h); (a') 2% glycine-[¹³C_a, ¹⁵N] in n. a. glycine (exp. time ~47 h); (b) 1% glycine-[¹³C', ¹⁵N] in glycine-[¹²C₂, ¹⁴N] (exp. time ~90 h); (b') 2% glycine-[¹³C', ¹⁵N] in n. a. glycine (exp. time ~40 hours).

neutron scattering (Table II). When r_{NMR} is determined from positions of the "singular" peaks in the powder spectrum $(\theta \approx \pi/2)$ one can introduce the deviations $r=r_0+\Delta$, $|\Delta| \ll r_0$, and $\theta \equiv (\pi/2) - \delta$, $|\delta| \ll 1$, and estimate the effect of atomic vibrations on r_{NMR} as

$$r_{\rm NMR} \equiv \left(\frac{\gamma_I \gamma_S \hbar}{\omega_D}\right)^{1/3} = r_0 \left(1 + 6\frac{\langle (\Delta)^2 \rangle}{r_0^2} - 3\langle \delta^2 \rangle\right)^{-1/3}$$
$$\approx r_0 \left(1 - 2\frac{\langle \Delta^2 \rangle}{r_0^2} + \langle \delta^2 \rangle\right), \tag{3}$$

where r_0 is the equilibrium distance between nuclei. Alternatively, the equilibrium distance r_0 can be calculated from r_{NMR} ,

$$r_0 \simeq r_{\rm NMR} (1 - \langle \delta^2 \rangle) + \frac{2\langle \Delta^2 \rangle}{r_{\rm NMR}},\tag{4}$$

when the variances of Δ and δ are known. C_{α} -N bond is rigid, and it is reasonable to assume that C_{α} -N stretching is close to one of the crystal normal modes. Then, $\langle \Delta^2 \rangle$ can be estimated from temperature and frequency of the corresponding harmonic oscillator. For an oscillator, $(1/2)m\omega^2 \langle \Delta^2 \rangle$ = $(1/2)E_{tot}$, where *m* is the reduced mass, ω is the frequency, and E_{tot} is the total energy. The average energy of harmonic

TABLE II. Comparison of experimental results for C_{α} -N distance in α -glycine.

Solid-state NMR	This work NMR, ${}^{13}C-{}^{15}N$, Refs. 16 and 29	1.496±0.002 Å 1.505(-0.001,0.002) Å
	10% glycine-[${}^{13}C_{\alpha}$, ${}^{15}N$] in n. a. glycine	
	NMR, ${}^{13}C-{}^{14}N$, single crystal, Ref. 24 9% glycine- ${}^{13}C_2$ in n. a. glycine	1.509±0.009 Å
X ray	Neutron scattering I, Ref. 26	1.476±0.001 Å
and	Neutron scattering II, Ref. 27	1.475±0.001 Å
Neutron	X-ray- diffraction, Ref. 25	1.474±0.005 Å

oscillator is $\overline{E} = \hbar \omega [(1/2) + (1/\exp(\hbar \omega/k_B T) - 1)]$ and, therefore,

$$\langle \Delta^2 \rangle = \frac{\bar{E}}{m\omega^2} = \frac{\hbar}{m\omega} \left[\frac{1}{2} + \frac{1}{\exp(\hbar\omega/k_B T) - 1} \right].$$
(5)

At $\hbar \omega \gg k_B T \langle \Delta^2 \rangle \cong (\hbar/m\omega)[(1/2) + \exp(-\hbar\omega/k_B T)]$, and at $\hbar \omega \ll k_B T \langle \Delta^2 \rangle \cong k_B T / m \omega^2$. From the IR spectrum of glycine, it is known²⁸ that the line at 1032 cm⁻¹ represents the C_{α} -N stretching. Using Eq. (5) one can find that C_{α} -N stretching gives $\langle \Delta^2 \rangle = 2.378 \times 10^{-3} \text{ Å}^2$. This is a small quantity and produces only 0.3% decrease of $r_{\rm NMR}$ compared to r_0 . Since $\hbar \omega > k_B T$ for this vibration, the difference is mostly contributed by a quantum uncertainty in r. It is more difficult to estimate $\langle \delta^2 \rangle$, which increases $r_{\rm NMR}$ compared to r_0 , because it may have contributions from many low-frequency thermally excited modes. According to molecular-dynamics simulation,²⁹ the average variance of the Euler angles for molecular librations of the C_{α} -N bond in glycine is $\langle \delta^2 \rangle$ =0.012 73 rad². It would make $r_{\rm NMR}$ longer than r_0 by about 1% or 0.02 Å, which is comparable to the differences between the results obtained with NMR and other techniques (Table II).

VII. ¹⁵N DETECTION

With further improved sensitivity it may become advantageous to use ¹⁵N detection. Lower γ of the ¹⁵N nucleus results in decreased broadening factors, and lower natural abundance makes the results obtained in natural-abundance matrices almost as good as in isotopically depleted matrices. The scheme of experiment is the same as in Fig. 1 except that ¹³C is replaced by ¹⁵N and vice versa. The adiabatic ¹⁵N pulse had 30 ms duration, 30 kHz sweeping range, and 1.3 kHz amplitude. The dipolar powder spectra for the four glycine samples discussed above are presented in Fig. 6. Because of higher noise level the powder patterns are not distinguishable for the spectra in Figs. 6(a) and 6(a'). At the same time, significantly reduced line broadening makes the "singular" peaks clearly visible well above the noise level. We believe that the data obtained with ¹⁵N detection and shown in parentheses in Table I provide more accurate results than ¹³C detection. However, we did not attempt to use the ¹⁵N data for reducing the error margins of the presented $r_{\rm NMR}$ values because of high noise level of the spectra in Fig. 6.

VIII. CONCLUSION

The major factor limiting an accuracy of NMR distance measurements in our experiments was line broadening, resulting from nonperfect protons decoupling. We used a conventional probe for liquids and the available protons decoupling power was only 50 kHz. We think that the accuracy can be considerably increased with using a customized probe, which will have optimized sensitivity for a directly detected nucleus and, at the same time, will allow highpower protons decoupling. In fact, the accuracy of the NMR distance measurement can be so high that such data can be used for refining structures obtained with x ray or neutron scattering. However, molecular vibrations affect differently the results of different methods, and more work on quantifying the vibrational contributions is needed before different techniques can complement each other.

ACKNOWLEDGMENTS

The authors thank Professor C. Jaroniec and Professor M. Tubergen for helpful discussions and Dr. M. Gangoda for technical assistance. The work was supported by Kent State University and Ohio Board of Regents.

- ¹K. Wüthrich, *NMR of Proteins and Nucleic Acids* (Wiley, New York, 1986).
- ² J. W. Emsley and J. C. Lindon, *NMR Spectroscopy Using Liquid Crystal Solvents* (Pergamon, Elmsford, New York, 1975).
- ³N. Tjandra and A. Bax, Science **278**, 1111 (1997).
- ⁴D. Sandström and M. H. Levitt, J. Am. Chem. Soc. **118**, 6966 (1996).
- ⁵M. Hong, A. Pines, and S. Caldarelli, J. Phys. Chem. **100**, 14815 (1996).
- ⁶B. M. Fung, K. Ermolaev, and Y. Yu, J. Magn. Reson. 138, 28 (1999).
- ⁷J. S. Waugh, Proc. Natl. Acad. Sci. U.S.A. **73**, 1394 (1976).
- ⁸C. H. Wu, A. Ramamoorthy, and S. J. Opella, J. Magn. Reson., Ser. A 109, 270 (1994).
- ⁹ A. Ramamoorthy, Y. Wei, and D.-K. Lee, Annu. Rep. NMR Spectrosc. **52**, 1 (2004).
- ¹⁰D. P. Raleigh, M. H. Levitt, and R. G. Griffin, Chem. Phys. Lett. **146**, 71 (1988).
- ¹¹ T. Fujiwara, P. Khandelwal, and H. Akutsu, J. Magn. Reson. **145**, 73 (2000).
- ¹²T. Gullion and J. Schaefer, J. Magn. Reson. **81**, 196–200 (1989).
- ¹³ M. Carravetta, M. Edén, O. G. Johannessen, H. Luthman, P. J. E. Verdegem, J. Lugtenburg, A. Sebald, and M. H. Levitt, J. Am. Chem. Soc. **123**, 10628 (2001).
- ¹⁴C. S. Yannoni and R. D. Kendrick, J. Chem. Phys. **74**, 747 (1981).
- ¹⁵M. Engelsberg and C. S. Yannoni, J. Magn. Reson. **88**, 393 (1990).
- ¹⁶Y. Ishii and T. Terao, J. Magn. Reson., Ser. A **115**, 116 (1995).
- ¹⁷J.-S. Lee and A. K. Khitrin, J. Magn. Reson. **177**, 152 (2005).
- ¹⁸A. E. Bennett, C. M. Rienstra, M. Auger, K. V. Lakshmi, and R. G. Griffin, J. Chem. Phys. **103**, 6951 (1995).
- ¹⁹B. M. Fung, A. K. Khitrin, and K. Ermolaev, J. Magn. Reson. **142**, 97 (2000).
- ²⁰A. K. Khitrin and B. M. Fung, J. Chem. Phys. **112**, 2392 (2000).
- ²¹ A. K. Khitrin, T. Fujiwara, and H. Akutsu, J. Magn. Reson. **162**, 46 (2003).
- ²² R. Freeman and H. D. W. Hill, J. Chem. Phys. **54**, 301 (1971).
- ²³ R. Freeman, Spin Choreography (Oxford University Press, Oxford, 1998).
- ²⁴R. A. Haberkorn, R. E. Stark, H. van Willigen, and R. G. Griffin, J. Am. Chem. Soc. **103**, 2534 (1981).
- ²⁵ R. E. Marsh, Acta Crystallogr. **11**, 654 (1958).
- ²⁶P.-G. Jönsson and Å. Kvick, Acta Crystallogr., Sect. B: Struct. Crystallogr. Cryst. Chem. **B28**, 1827 (1972).
- ²⁷L. F. Power, K. E. Turner, and F. H. Moore, Acta Crystallogr., Sect. B: Struct. Crystallogr. Cryst. Chem. **B32**, 11 (1976).
- ²⁸C. L. Thaper, B. A. Dasannacharya, P. S. Goyal, R. Chakravarthy, and J. Tomkinson, Physica B **174**, 251 (1991).
- ²⁹ Y. Ishii, T. Terao, and S. Hayashi, J. Chem. Phys. 107, 2760 (1997).